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Article in *Journal of Advances in Microbiology* · October 2022

DOI: 10.9734/JAMB/2022/v22i11677

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***In-vitro* Antibacterial Activity of *Allium sativum* on Common Food Poisoning Bacteria**

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Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/JAMB/2022/v22i11677

Open Peer Review History:

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: <https://www.sdiarticle5.com/review-history/92240>

Original Research Article

Received 20 July 2022
Accepted 26 September 2022
Published 05 October 2022

ABSTRACT

Background: The use of herbal medicines for food poisoning in inadequate resource countries has greatly increased with the limited establishment of antibacterial activity. This study aimed at establishing the antibacterial activity of the aqueous *Allium sativum* extract against *Staphylococcus aureus*, *Escherichia coli* and *Salmonella typhimurium*.

Methods: *Allium sativum* L. (Amaryllidaceae) was purchased from a farmer in Kasese District and was authenticated. The material was then extracted using distilled water, filtered and lyophilized. Evaluation of the phytochemicals in the extract was done using standard procedures. High-Performance Liquid Chromatography (HPLC) fingerprint of the extract was conducted to monitor the reproducibility of the extract. The antibacterial activity of the aqueous garlic extract against the organisms above was evaluated and expressed as Minimum Inhibitory Concentration (MIC) and Minimum Bactericidal Concentration (MBC).

Results: Phytochemical screening of the garlic extract showed the presence of amino acids, alkaloids, glycosides and saponins, flavonoids, steroids and terpenoids. HPLC fingerprints showed

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39 and 98 peaks at 220nm and 254nm wavelengths respectively and the presence of various phytochemical compounds in *A. sativum*. The extract showed antibacterial activity against *S. aureus*, *E. coli* and *S. typhi*.

Conclusion: The aqueous garlic extract exhibited antibacterial activity against tested common food poisoning bacteria, with a high potency against gram-positive bacteria.

Keywords: *Allium sativum*; antibacterial; food poisoning; *Staphylococcus aureus*; *Escherichia coli*; *Salmonella typhimurium*.

1. INTRODUCTION

Allium sativum L. (garlic) is a small, ubiquitous species in the family Amaryllidaceae, Genus *Allium* and commonly used food spice for food processing [1]. In addition to being an aromatic herbaceous plant that is consumed worldwide as food, *A. sativum* is also used as a traditional remedy for various diseases such as food poisoning, bronchitis, intestinal worms, etc. [2]. Among these, food poisoning has increasingly become a global health challenge, which requires interventions, stemming up right from our local communities [3]. According to the World Health Organization, an estimated 600 million; almost 1 in 10 people in the world fall ill after eating contaminated food and 420,000 die every year, with the WHO African and South-East Asia Regions having the highest morbidity and mortality rates, including children under the age of 5 years [4]. Food poisoning is a group of illnesses acquired by consumption of foods contaminated with infective organisms and/or their toxins, chemical contaminants, whether metallic or organic [5]. Food contamination occurs from food processing, preparation, and storage leading to sickness; hospitalizations or eventually death otherwise called foodborne diseases [6]. The infective organisms that contaminate food include bacteria, viruses and parasites, which cause more than 200 diseases like diarrhea, cancers, etc. [4,6]. Antibiotics such as ciprofloxacin, azithromycin, rifamycin, etc. are some of the common treatments usually recommended for food poisoning infections caused by bacteria [7-9]. However, their prolonged administration and abuse have been linked to the emergence and spread of resistant bacteria, rendering the treatment of infectious diseases ineffective [10]. This increasing microbial resistance to conventional drugs has revived the interest in plants with antibacterial activities. Available literature shows that garlic has been widely known for its potency of dietary and medicinal use to cure various diseases, including infectious diseases for centuries [11]. Garlic has also been used as a home-based

prophylactic and symptom management agent in the recent COVID-19 emergence, it was reported to have therapeutic efficacy against multiple symptoms observed in advanced COVID-19 patients [12]. The antibacterial activity of *A. sativum* has been mainly attributed to three phytochemical families, fructans (mainly inulin), phenolic compounds (flavonoids), and organosulfur compounds (OSCs) [13]. It is also known that the most sensitive bacterium to garlic is the deadly *Bacillus anthracis* which causes the disease anthrax [14]. Some studies have also shown garlic to have antibacterial activity against bacteria [15,16]. However, the antibacterial activity of garlic on common food poisoning bacteria has not been extensively studied and this was investigated in this study.

2. METHODOLOGY

2.1 Study Design

It was a laboratory-based study in which *A. sativum* active ingredients were extracted using distilled water, and the presence phytochemicals in the extract were determined. The aqueous garlic extract obtained was evaluated for antibacterial activity against *Staphylococcus aureus* (Cat. 25923, ATCCTM), *Escherichia coli* (Cat. 25922, ATCCTM) and *Salmonella typhimurium* (Cat. 14028, ATCCTM).

2.2 Study Area

Allium sativum L. (family Amaryllidaceae) was obtained from a local garden in Mubuku village, Kasese district, Uganda and the study was conducted in Pharmaceutical Chemistry/Analysis Laboratory of Mbarara University of Science and Technology, Mbarara (MUST), where garlic extraction and evaluation of the phytochemicals in the extract was done, and the Microbiology Laboratory of MUST, where the determination of the antibacterial activity of the garlic extract against standard food poisoning bacteria was done.

2.3 Study Population

Allium sativum L. was used in this study. The purposive sampling method was used to obtain the garlic from the local farmer's garden in Mubuku village, Kasese district, before it was authenticated by Dr. Olet Eunice, a botanist in the Biology Department of the Faculty of Science, MUST.

2.4 Preparation of Garlic Extract

The sterile aqueous extract of garlic was obtained using a method by Benmeziane *et al.* [17], where the cloves on fresh bulbs garlic obtained were peeled using a kitchen knife and washed thoroughly under running tap water to obtain the edible portion. The garlic extract was prepared immediately from 1 kg to avoid degradation of allicin [18], by cutting into pieces [19] and crushed to release allinase under aseptic conditions before it was mixed with distilled water to allow alliin to be converted to allicin by enzyme allinase [20]. The homogenate was recovered by filtration through a double-layered sterile fine mesh cloth into an Eppendorf tube [21]. An extract sample of 800 ml was then lyophilized using a freeze-dryer for 3 days. The obtained garlic powder was stored at 4 °C till use [22].

2.5 Phytochemical Screening of Garlic Extract

Following methods were used to determine presence of phytochemicals: quinones (Borntrager's test) [23], alkaloids (Dragendorff's test) [24], glycosides (Benedict's test) [25], phenols (FeCl_3) [26], tannins (Folin-Ciocalteu method) [27], flavonoids (Ammonia test) [28], saponins (Frothing test) [29], steroids (Chloroform and concentrated H_2SO_4 meth) [30], terpenoids (Chloroform and concentrated H_2SO_4) [26] and thiosulphates ethyl (p-hydroxybenzoate) [31], amino acids (ninhydrin test) [32].

2.6 High Performance Liquid Chromatography Apparatus for Fingerprint

The high-performance liquid chromatography (HPLC) fingerprint was performed on a UFLC Shimadzu Prominence Model HPLC system (Tokyo, Japan) at the Analytical and Pharmaceutical Laboratory, Mbarara University of Science and Technology, Uganda, comprising

an LC-20AD pump, a Phenomenex Luna C_{18} column (250×4.6 mm. 5 μm), temperature-controlled sample trays, an online degasser DGU-20A5R and an ultraviolet (UV) detector. A reversed-phase HPLC assay was carried out using a binary isocratic elution with a flow rate of 1.0 ml/min at a column temperature of 25 °C, a mobile phase of methanol/acetonitrile /0.01% trifluoroacetic acid (6:1:3). The injection volume was 10 μL at detection wavelengths of 220 and 254 nm while the acquisition time was 40 min.

2.7 Evaluation of Antibacterial Activity

Antibacterial activities of the sterile aqueous garlic extracts (at concentrations of 1000, 500, 250, 125, 62.5 mg/ml), 100-folds diluted ciprofloxacin (positive control) and distilled water (negative control) were evaluated using agar well diffusion method on Mueller Hinton Agar (MHA) (Cat. R454084, Thermofisher). The Zones of inhibition were measured using a ruler. Single colonies on agar plates were used to prepare the bacterial suspension with a turbidity of 0.5 McFarland (equal to 1.5×10^8 colony-forming units (CFU)/ml) [17].

2.8 Determination of MIC and MBC

The Minimum Inhibitory Concentration (MIC) and Minimum Bactericidal Concentration (MBC) of aqueous garlic extract were determined using following the Alirezaei's method [33] and Shetty's method [34] respectively.

2.8.1 Determination of Minimum Inhibitory Concentration (MIC)

The MHA plates were inoculated with standard bacterial strain under aseptic conditions and wells of 8 mm diameter were made using a sterile cork borer. *A. Sativum* extracts were prepared in a series of decreasing concentrations. One hundred (100) μL of extracted garlic were introduced into the wells using automatic microliter pipettes, and all plates were incubated at 37°C for 24 hours. The sensitivity of *S. typhimurium*, *E. coli* and *S. aureus* was determined by measuring the diameter of the zone of inhibition.

2.8.2 Determination of Minimum Bactericidal Concentration (MBC)

Brain Heart Infusion (BHI) broth (Cat.17580547, Oxoid, Thermofisher, China) of 900 μL was poured into sterile tubes. Then, each tube was

inoculated with 100 μ L of the standardized microbial suspension (0.5 Mc-Farland). After mixing, the inoculated tubes were incubated. The negative control used was distilled water while the tubes were sub-cultured on MHA and incubated for 24 hours at 37°C.

2.9 Data Analysis

All quantitative data were expressed as mean $\pm$ standard error of mean (SEM) while the variation in a set of data was analyzed through the one-way Analysis of Variance. The difference among the means was considered at 95% confidence level using the post-hoc method of Tukey's Multiple Comparison Test through GraphPad Prism9.

3. RESULTS

3.1 Phytochemical Screening and Antibacterial Activity of the Extract

The phytochemical analysis of the extract of *Allium sativum* showed presence of amino acids, alkaloids, glycosides, saponins, flavonoids, steroids and terpenoids. The extract showed antibacterial activity against all the tested microorganisms that is *Staphylococcus aureus*, *Escherichia coli* and *Salmonella typhimurium*. In order to ensure the reproducibility of the extract

for quality control purposes, the HPLC fingerprint conducted using both 220 and 254nm showed 39 and 98 peaks, respectively.

3.2 High Performance Liquid Chromatography Fingerprint

The HPLC fingerprints obtained at wavelengths of 220 and 254nm as shown in Figs. 1 and 2 below showed chromatograms with 39 peaks (at 220nm) and 98 peaks (at 254nm). The peaks seen in the fingerprints correspond to the phytochemical compounds in the *Allium sativum*.

3.3 Antibacterial Activity of *A. sativum* Extract

3.3.1 Agar-well diffusion

The garlic extract had the largest zones of inhibition on *S. aureus*, followed by *S. typhimurium* and *E. coli* being the least across all concentrations (Figs. 3 and 4). The garlic extract was 2-fold more effective on *S. aureus* than *S. typhimurium* and *E. coli* (Figs. 3 and 4). The garlic extract was also more effective on *S. aureus* than the standard drug, ciprofloxacin. However, ciprofloxacin was more effective on *E. coli* and *S. typhimurium* than the garlic extract (Figs. 3 and 4).

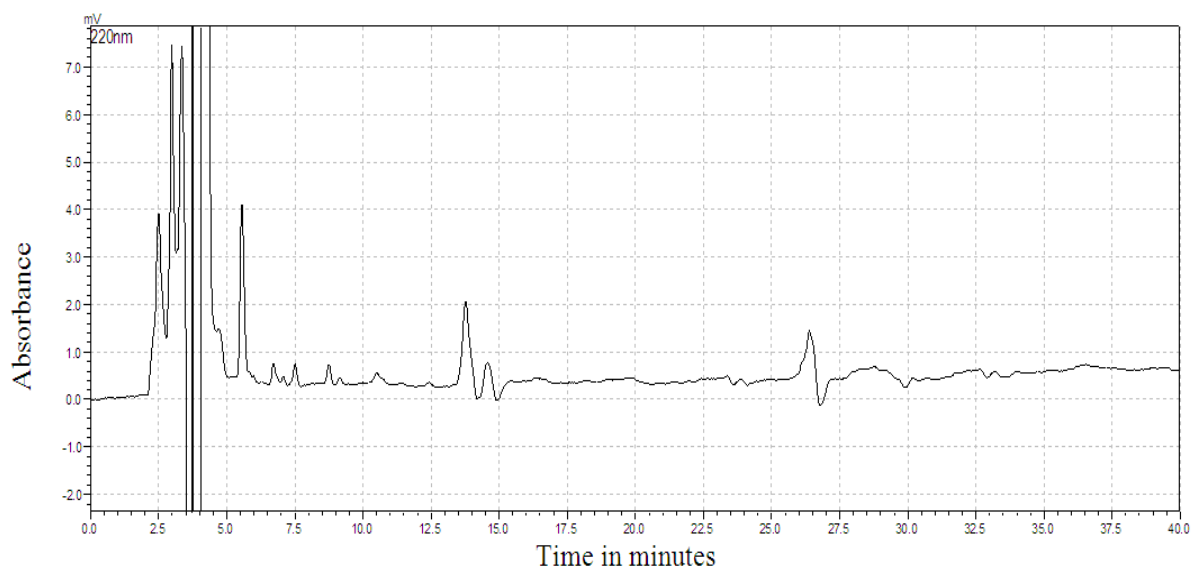


Fig. 1. HPLC fingerprint of *A. sativum* extract at 220 nm wavelength

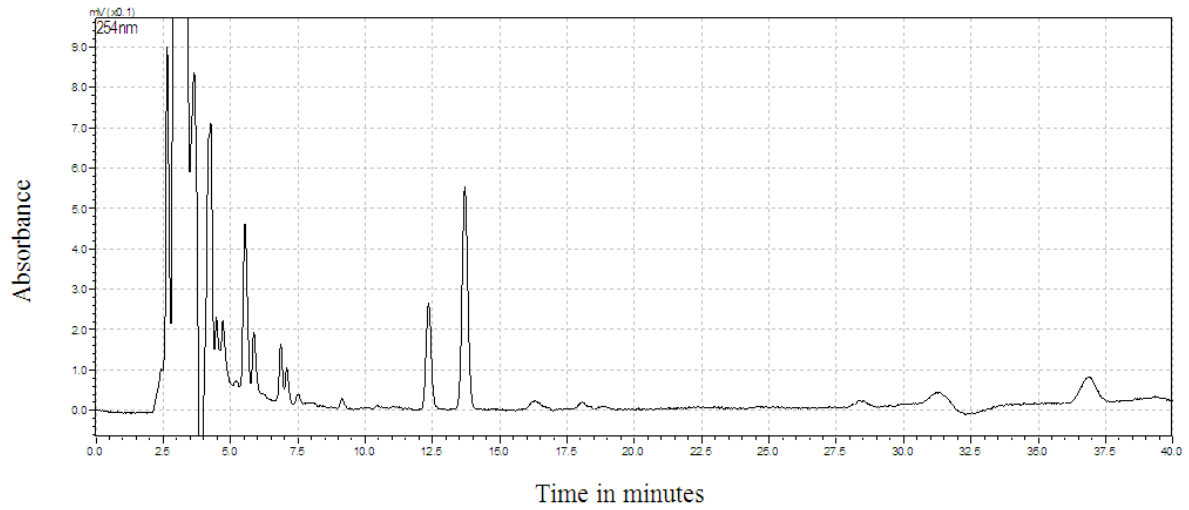


Fig. 2. HPLC fingerprint of *A. sativum* extract at 254 nm wavelength

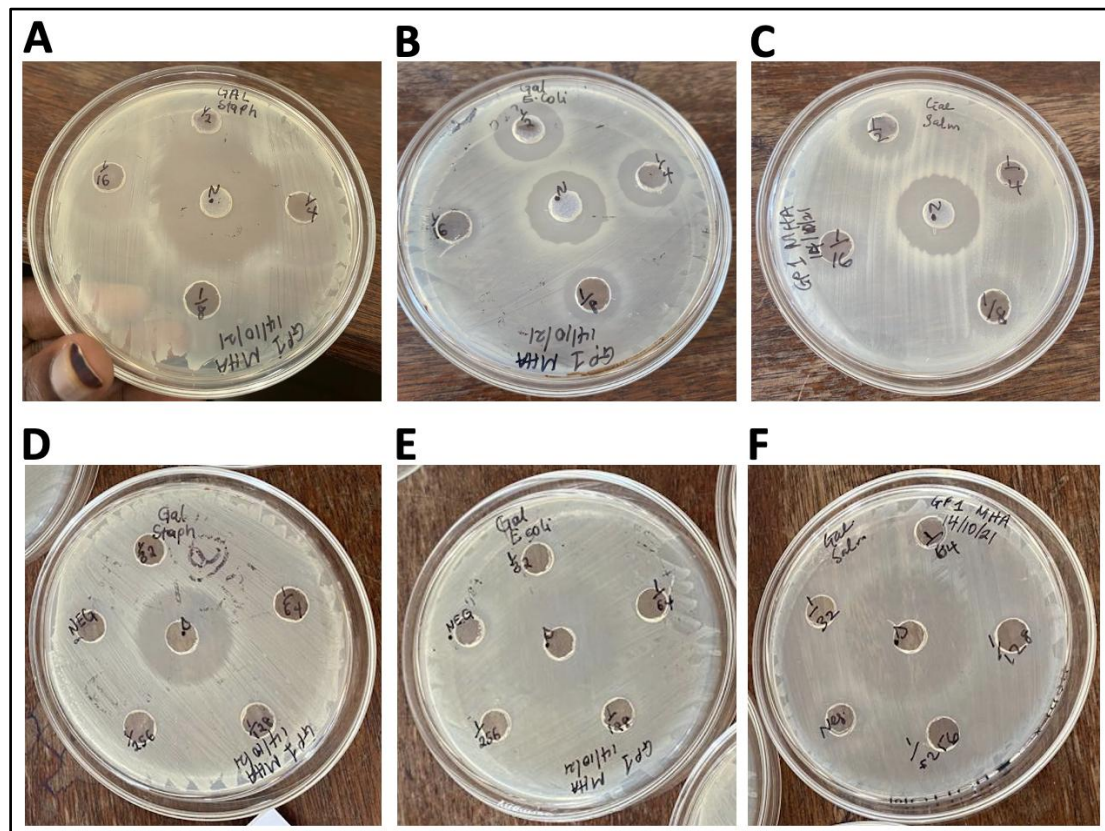


Fig. 3. Agar-well diffusion images (A, B and C show the effects of garlic extract on *S. aureus*, *E. coli* and *S. typhimurium* respectively; E, F and G show the effects of controls on *S. aureus*, *E. coli* and *S. typhimurium* respectively where label D denotes positive control, ciprofloxacin and label NEG denotes negative control)

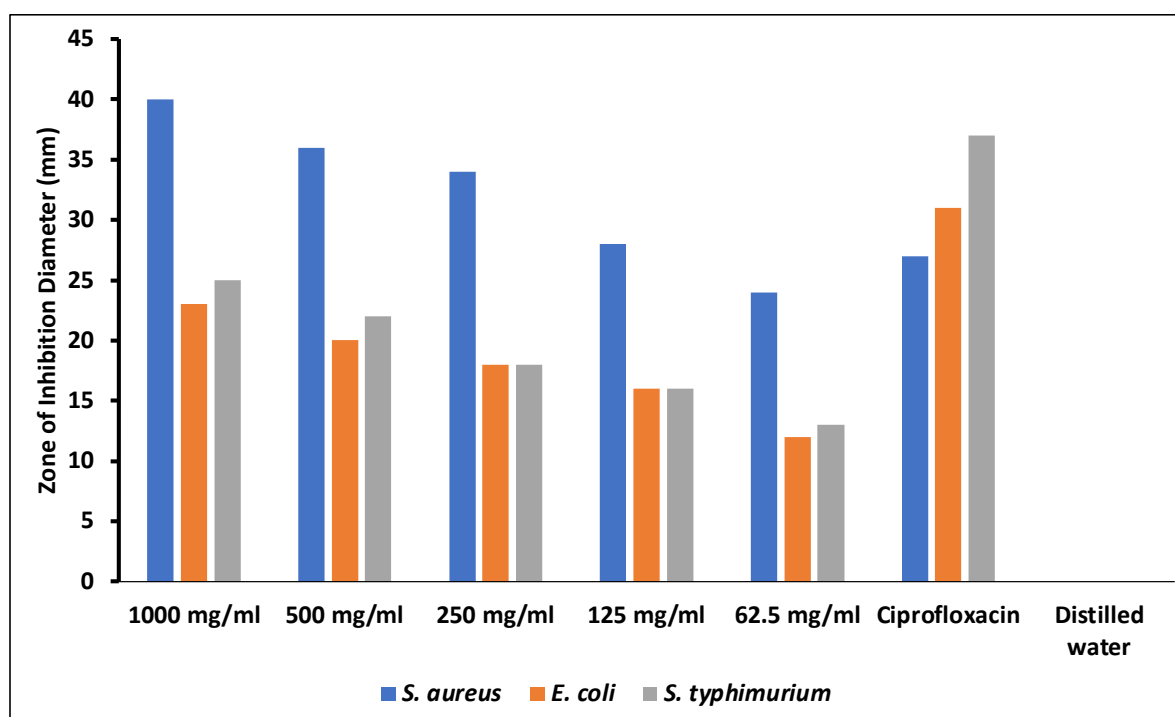


Fig. 4. Zone of inhibition diameters after 24 hours at different concentrations of garlic extract, 100-folds diluted intravenous ciprofloxacin (positive control) and distilled water (negative control)

Table 1. MIC values of the aqueous *Allium sativum* extract

Organisms	Average MIC (mg/ml)
<i>Staphylococcus aureus</i> (ATCC 25923)	31.25
<i>Escherichia coli</i> (ATCC 25922)	62.5
<i>Salmonella typhimurium</i> (ATCC 14028)	62.5

Table 2. MBC values of the aqueous *Allium sativum* extract.

Organisms	Mean MBC (mg/ml)
<i>Staphylococcus aureus</i> (ATCC 25923)	125
<i>Escherichia coli</i> (ATCC 25922)	250
<i>Salmonella typhimurium</i> (ATCC 14028)	

3.3.2 Minimum Inhibitory Concentrations (MIC) & Minimum Bactericidal Concentrations (MBC)

The *Allium sativum* extract showed a profound antibacterial activity against all the tested organisms with lowest Minimum Inhibitory Concentration value of 31.25 mg/ml against *Staphylococcus aureus*, and highest (62.5 mg/ml) against both *Escherichia coli* and *Salmonella typhimurium* (Table 1).

Also, the Minimum Bactericidal Concentration of the aqueous *Allium sativum* extract on *S. aureus* and *S. typhimurium* had the highest MBC of 125

mg/ml while on *E. coli*, it had the least MBC value of 250 mg/ml (Table 2).

4. DISCUSSION

In previous study, it was reported that the antibacterial activity of garlic is attributed to its phytochemical constituents such as fructans (mainly inulin), phenolic compounds (flavonoids), and organosulfur compounds (OSCs) [35]. Likewise in this study, phytochemical screening showed presence of amino acids, alkaloids, glycosides, saponins, flavonoids, steroids and terpenoids and these could be responsible for the observed antibacterial activity of the A.

sativum extract against *Staphylococcus aureus*, *Escherichia coli* and *Salmonella typhimurium*, which is in agreement with a study by Njue, et al 2014 (Njue, Kanja [36].

The peaks observed from the HPLC fingerprints in Figs. 1 and 2 correspond to the phytochemical compounds in the *Allium sativum* extract which is comparable to a study by Zhao, Li [37]. Zhao, Li and colleagues standardized the HPLC fingerprint by running standard solutions with their extracts. Therefore, they corresponded the peaks obtained from the extracts to those obtained from the standards and were able to specifically identify the phytochemical compounds, unlike in this study where standards were not used. The peak areas are equal to the peak height times width at half height. The larger the area, the more the amount of the phytochemical present in the extract. Peaks obtained at the same retention time at the two wavelengths represent the same compound. Some peaks are observed at one wavelength and not the other as shown in Figs. 1 and 2. Presence of a peak at retention times between 25 and 27.5 minutes only under a wavelength of 220nm indicated presence of a compound that is visualized at 220nm wavelength and not at 254nm. It is therefore always necessary to run the HPLC of the garlic extract at two different wavelengths in order to identify all the phytochemical compounds in the extract.

The antibacterial activity of the aqueous *Allium sativum* extract on three standard organisms studied varies with the type and composition of the extract, concentration used, types of microorganism, pH value and temperature of the environment [38]. The results of this study indicated that the aqueous extract of garlic had an antibacterial activity on the tested organisms. The extract was more active at high concentration and reduced at very low concentrations which is in accordance with a study by De Moura Oliveira, Santos-Mendonça [39]. Thus, the inhibition of bacterial growth by the aqueous garlic extract showed a dose dependent activity as reported by Shams et al. [40]. The susceptibility of the tested organisms to the garlic extract was expressed as MIC and MBC.

The MIC value of the garlic extract was lowest (31.25 mg/ml) against *Staphylococcus aureus*, a gram-positive bacterium and highest (62.5 mg/ml) against both *Escherichia coli* and *Salmonella typhimurium*, gram negative bacteria. The extract in our study is thus more potent

against gram positive bacteria than gram negative bacteria, as earlier reported by Nakamoto and Abubakar [41,42]. These observations are likely to be a result of the differences in the cell wall structure between gram negatives and gram positives, with gram negative outer membrane acting as a barrier to many environmental substances, including antibiotics [43]. The outer membrane, a characteristic of gram-negative bacteria like *Escherichia coli* and *Salmonella typhimurium* functions as a molecular sieve through which molecules with molecular mass >600–1000 Da cannot penetrate [44].

Additionally, the MIC values obtained in our study indicate that the extract is bacteriostatic against *Staphylococcus aureus* at an MIC of 31.25mg/ml, which slightly differs from a study by Simeon and Falilat [45] where a bacteriostatic activity of the garlic extract against *Staphylococcus aureus* was obtained at an MIC of 25 mg/ml.

On the other hand, a bacteriostatic activity of the extract against *Escherichia coli* obtained at the MIC of 62.5 mg/ml varied from the report of Simeon and Falilat [45] which was obtained at a slightly higher MIC value of 100 mg/ml while in *Salmonella typhimurium*, the bacteriostatic activity of the extract obtained at the MIC of 62.5mg/ml was slightly higher than that reported by Simeon and Falilat [45] which was obtained at the MIC of 25 mg/ml. The variation in the results obtained could be attributed to the difference in the garlic source, which directly affects the quantity of phytochemical composition responsible for the antibacterial activity of garlic [35].

The MBC values obtained in this study were 125 mg/ml for *S. aureus* and 250 mg/ml for *E. coli*, which indicates that the extract was bactericidal against the organisms at those concentrations. This varied from a study by Mohammad et al. (2020) [46] who obtained the bactericidal activity at an MBC of 100mg/ml for *S. aureus* and an MBC of 6.25 mg/ml for *E. coli* whereas for *S. typhimurium*, the bactericidal activity obtained at the MBC of 125 mg/ml was slightly higher than that in a study by Simeon and Falilat [45] which was obtained at the MBC of 50 mg/ml.

The quantification of active components using HPLC could not be done, since a standard was not used. Further studies to quantify the active compounds of *Allium sativum* and *in-vivo* studies to ascertain their bioavailability in body fluids,

which is key to the efficacy of drugs, need to be conducted.

5. CONCLUSION

The study showed that the aqueous garlic extract from Mubuku in Kasese District exhibited antibacterial activity against tested organisms, with a high potency against gram positive bacteria. The extract contained bioactive phytochemicals which could be responsible for its antibacterial activity. Gram-positive bacteria (*S. aureus*) more sensitive to garlic extracts than the gram-negative bacteria (*E. coli* and *S. typhimurium*). The high susceptibility of the tested bacterial isolates to the garlic extracts used in this study, gives evidence that the extracts contain compounds that can serve as suitable antibiotics against *S. aureus*, *E. coli* and *S. typhimurium*. However, further studies need to be conducted to assess the safety of the extracts especially their cytotoxicity. The antimicrobial effects of the garlic extracts on other bacterial species that have shown resistance to most of the antibiotics routinely used in treatment of bacterial infections should also be studied.

ACKNOWLEDGEMENTS

This study was funded by Mbarara University of Science and Technology, Uganda. We acknowledge the laboratory and entire staff, at the department of medical laboratory science, faculty of medicine whose efforts were invaluable.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Kshirsagar MM, et al. Antibacterial activity of garlic extract on cariogenic bacteria: An *In vitro* study. *Ayu*. 2018;39(3):165-168.
2. Batiha GES, Beshbishy AM, Wasef LG. Chemical Constituents and Pharmacological Activities of Garlic (*Allium sativum* L.): A Review; 2020.
3. Mohammad AM, et al. Food poisoning and intoxication: A global leading concern for human health, in Food safety and preservation. 2018;307-352.
4. WHO, Food Safety; 2020.
5. Bintsis T. Foodborne pathogens. *AIMS Microbiology*. 2017;3(3):529.
6. AL-Mamun M, et al. Chapter 11-Food poisoning and intoxication: A global leading concern for human health. *Food Safety & Preservation* (Edited by Grumezescu AM & Holban AM). Academic Press, United States. 2018;307-52.
7. Wong MHY, et al. Characterization of *Salmonella* food isolates with concurrent resistance to ceftriaxone and ciprofloxacin. *Foodborne Pathogens and Disease*. 2013;10(1):42-46.
8. Rogers AJ, Lee GHA, Boan P. Azithromycin for *Salmonella* infection: Don't presume it works. *The Medical Journal of Australia*. 2018;209(2):98.
9. Lin SW, Lin CJ, Yang JC. Rifamycin SV MMX for the treatment of traveler's diarrhea. *Expert Opinion on Pharmacotherapy*. 2017;18(12):1269-1277.
10. Morehead MS, Scarbrough C. Emergence of global antibiotic resistance. *Prim Care*. 2018;45(3):467-484.
11. Leyla Bayan, PHK, Ali Gorji. A review of potential therapeutic effects of garlic; 2013.
12. Thota SM, Balan V, Sivaramakrishnan V. Natural products as home-based prophylactic and symptom management agents in the setting of COVID-19. *Phytotherapy Research*. 2020;34(12):3148-3167.
13. Ramirez DA, Altamirano JC, Camargo AB. Multi-phytochemical determination of polar and non-polar garlic bioactive compounds in different food and nutraceutical preparations. *Food Chemistry*. 2021;337:127648.
14. Hawassa E, Sekota E. Anti-Bacterial effect of garlic (*Allium sativum*) against clinical isolates of *Staphylococcus aureus* and *Escherichia coli* from patients attending hawassa referral hospital, Ethiopia. *Journal of Infectious Diseases*. 2016;2(2):18.
15. Nejad ASM, et al. Antibacterial effect of garlic aqueous extract on *Staphylococcus aureus* in hamburger. *Jundishapur Journal of Microbiology*. 2014;7(11).
16. Kshirsagar MM, et al. Antibacterial activity of garlic extract on cariogenic bacteria: An *In vitro* study. *Ayu*. 2018;39(3):165.
17. Benmeziane F, et al. Evaluation of antibacterial activity of aqueous extract and essential oil from garlic against some pathogenic bacteria. *International Food Research Journal*. 2018;25(2).

18. Farías-Campomanes AM, et al. Allicin-rich extract obtained from garlic by pressurized liquid extraction: Quantitative determination of allicin in garlic samples. *Food and Public Health*. 2014;4(6):272-278.
19. Ratti C, et al. Drying of garlic (*Allium sativum*) and its effect on allicin retention. *Drying Technology*. 2007;25(2):349-356.
20. Borlinghaus J, et al. Allicin: Chemistry and biological properties. *Molecules*. 2014;19(8):12591-12618.
21. Prati P, et al. Evaluation of allicin stability in processed garlic of different cultivars. *Food Science and Technology*. 2014;34(3):623-628.
22. Bajac J, et al. Extraction of different garlic varieties (*Allium sativum* L.)-Determination of organosulfur compounds and microbiological activity. In *International Congress Food Technology. Quality and Safety*; 2018.
23. Houghton P, Raman A. *Laboratory handbook for the fractionation of natural extracts*. Springer Science & Business Media; 2012.
24. Sarma D, Koul S, Khosa R. Alkaloids from *Tinospora cordifolia* miers. *J Pharm Sci Res*. 2009;1:26-7.
25. Kumar A, et al. Preliminary phytochemical analysis of leaf and bark (mixture) extract of *Ficus infectoria* plant. *The Pharma Innovation*. 2012;1(5, Part A):71.
26. Ahmed FA, Kabir H. Ethnomedicinal value, phytochemical composition and bioactivity of *Butea monosperma* (Lam.) Taub. *Jahangirnagar University Journal of Biological Sciences*. 2015;4(2): 19-29.
27. Pardede C, et al. Extraction of tannin from garlic skins by using microwave with ethanol as solvent. In *IOP Conference Series: Materials Science and Engineering*. IOP Publishing; 2020.
28. Sankhalkar S, Vernekar V. Quantitative and Qualitative analysis of Phenolic and Flavonoid content in *Moringa oleifera* Lam and *Ocimum tenuiflorum* L. *Pharmacognosy research*. 2016;8(1): 16.
29. Bhattacharjee A, Das A. Phytochemical screening of some Indian plants. *Quarterly Journal of Crude Drug Research*. 1969;9(3):1408-1412.
30. Compennolle F, Vanderhaeghe H, Nonclercq M. Detection and identification of trace steroids in prednisolone. *Journal of Pharmacy and Pharmacology*. 1972; 24(6):429-433.
31. Eagling D, Sterling S. A cholesterol-lowering extract from garlic. Australia: Rural Industries Research and Development Corporation; 2000.
32. Sun SW, et al. Efficiency improvements on ninhydrin method for amino acid quantification. *Journal of Food Composition and Analysis*. 2006;19(2-3):112-117.
33. Alirezai S, Godarzi H, Maheri A. Antimicrobial activity of aqueous garlic extract (*Allium sativum*) against *porphyromonas gingivalis*: An *In-Vitro* Study; 2019.
34. Shetty S, et al. An *in-vitro* evaluation of the efficacy of garlic extract as an antimicrobial agent on periodontal pathogens: A microbiological study. *Ayu*. 2013;34 (4):445-51.
35. Ramirez DA, Altamirano JC, Camargo AB. Multi-phytochemical determination of polar and non-polar garlic bioactive compounds in different food and nutraceutical preparations. *Food Chem*. 2021;337: 127648.
36. Njue L, et al. Efficacy of antimicrobial activity of garlic extracts on bacterial pathogens commonly found to contaminate meat. *East African Medical Journal*. 2014;91(12):442-448.
37. Zhao D, et al. HPLC Fingerprint characteristics of active materials of garlic and other *Allium* species. *Analytical Letters*. 2014;47(1):155-166.
38. Espinoza T, et al. Garlic (*Allium sativum* L) and Its beneficial properties for health: A review. *Agroindustrial Science*. 2020;10(1): 103-115.
39. De Moura Oliveira KA, et al. Aqueous garlic extract and microbiological quality of refrigerated poultry meat. *Journal of Food Processing and Preservation*. 2005;29(2):98-108.
40. Shams Ghahfarokhi M, et al. Inhibitory effects of aqueous onion and garlic extracts on growth and keratinase activity in *Trichophyton mentagrophytes*. *Iranian Biomedical Journal*. 2003;7(3):113-118.
41. EL-Mahmood MA. Efficacy of crude extracts of garlic (*Allium sativum* Linn.) against nosocomial *Escherichia coli*, *Staphylococcus aureus*, *Streptococcus pneumoniae* and *Pseudomonas aeruginosa*. *Journal of Medicinal Plants Research*. 2009;3(4):179-185.

42. Nakamoto M, et al. Antimicrobial properties of hydrophobic compounds in garlic: Allicin, vinylidithiin, ajoene and diallyl polysulfides. *Experimental and Therapeutic Medicine*. 2020;19(2):1550-1553.
43. Verma U, Kulshreshtha S, Khatri P. MDR *Pseudomonas aeruginosa* in nosocomial infection: burden in ICU and burn units of a tertiary care hospital. *IJCMAS*. 2018;7: 1267-74.
44. Gallucci MN, et al. Antimicrobial combined action of terpenes against the food-borne microorganisms *Escherichia coli*, *Staphylococcus aureus* and *Bacillus cereus*. *Flavour and Fragrance Journal*. 2009;24(6):348-354.
45. Simeon YT, Falilat UO. Antibacterial effects of aqueous extract of onion and garlic on some clinical bacterial isolates; 2018.
46. Mohammed MF, et al. Antibacterial activities of allium sativum (Garlic) extracts against *Staphylococcus aureus* and *Escherichia coli*. *Eur. J. Mol. Clin. Med*. 2020;7:526-534.

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